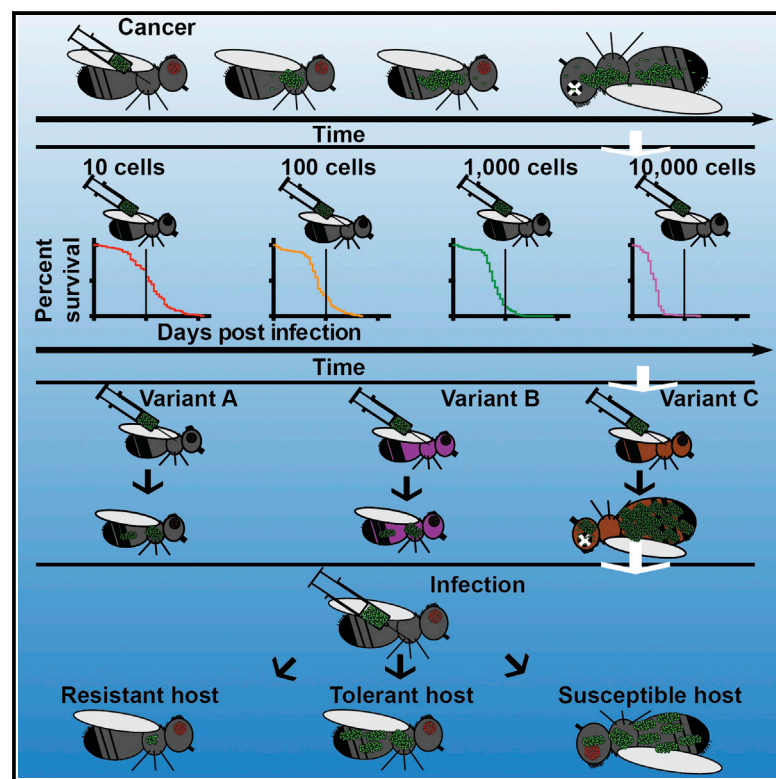


Cell Reports

Defining Resistance and Tolerance to Cancer

Graphical Abstract



Authors

Adler R. Dillman, David S. Schneider

Correspondence

dschneider@stanford.edu

In Brief

Dillman and Schneider develop a model to differentiate resistance and tolerance to cancer. They plot disease tolerance curves to cancer in wild-type flies and compare this to natural variants, identifying a line with reduced cancer resistance. Quantitation of these two traits opens an additional dimension for analysis of cancer biology.

Highlights

- We developed a framework for differentiating resistance and tolerance to cancer
- Cancer kills flies in a dose-dependent manner
- Natural variation affects cancer resistance
- We identified a fly line with reduced resistance to cancer



Defining Resistance and Tolerance to Cancer

Adler R. Dillman^{1,2} and David S. Schneider^{1,*}

¹Department of Microbiology and Immunology, Stanford University, Stanford, CA 94305, USA

²Present address: Department of Nematology, University of California, Riverside, Riverside, CA 92521, USA

*Correspondence: dschneider@stanford.edu

<http://dx.doi.org/10.1016/j.celrep.2015.09.052>

This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

SUMMARY

There are two ways to maintain fitness in the face of infection: resistance is a host's ability to reduce microbe load and disease tolerance is the ability of the host to endure the negative health effects of infection. Resistance and disease tolerance should be applicable to any insult to the host and have been explored in depth with regards to infection but have not been examined in the context of cancer. Here, we establish a framework for measuring and separating resistance and disease tolerance to cancer in *Drosophila melanogaster*. We plot a disease tolerance curve to cancer in wild-type flies and then compare this to natural variants, identifying a line with reduced cancer resistance. Quantitation of these two traits opens an additional dimension for analysis of cancer biology.

RESULTS AND DISCUSSION

Host immune defense strategies can be separated into the ability to control pathogen burden, called resistance, and the ability of the host to endure the negative health effects of infection, called disease tolerance. Disease tolerance is the dose-response curve relating host health to elicitor loads. While resistance is a heavily studied aspect of immune response, disease tolerance is less well understood. Originating in plant ecology studies (Caldwell et al., 1958; Schafer, 1971), the concept of disease tolerance was only recently introduced to animal immunity research (Ayres et al., 2008; Råberg et al., 2007). Distinguishing between resistance and disease tolerance is useful because they are fundamentally different strategies for surviving challenges. Applying the concepts of resistance and disease tolerance has improved our understanding of pathogenic infections (Iwasaki and Pillai, 2014; Medzhitov et al., 2012; Råberg, 2014; Vale et al., 2014) and should be applicable to any insult to host health, like cancer, not just infectious disease. We established a model to separate resistance and tolerance to cancer to understand the role of these immunological processes in cancer infections.

A System for Separating Resistance and Tolerance to Cancer

The model organism *Drosophila melanogaster* is useful for investigating both resistance and disease tolerance in infec-

tions because large numbers of animals can be infected with precise doses of pathogens and the growth of the pathogens and health of the host can be easily monitored (Ayres et al., 2008; Ayres and Schneider, 2009; Howick and Lazzaro, 2014; Rose et al., 2011; Rottschaefer and Lazzaro, 2012); we reasoned the fly would be suitable for studying resistance and tolerance to cancer. We used the *Drosophila* Oregon-R strain as an initial wild-type strain in our experiments. We chose to use a transplantable cancer model instead of an inducible one because it let us precisely regulate and measure input material (Ayres et al., 2008; Råberg et al., 2007; Regoes et al., 2014). We used the Ras^{v12}-H7 line of *Drosophila* hyperplastic cancer cells, which expresses an oncogenic form of *Ras*, has a UAS-GFP reporter, and has previously been shown to metastasize throughout the fly and lead to premature death (Simcox et al., 2008) (see [Experimental Procedures](#)). The hyperplastic cells were delivered in a manner similar to microbial pathogens; the cells were cultured in vitro, quantified, diluted, and injected into adult flies (Figures 1A and 1B). We used survival (median time to death) as a measure of disease progression and found that, similar to microbial infections, cancer kills in a dose-dependent manner, ranging from 8 to 21 days (Figure 1A) whereas wounding controls would live for 29–32 days. To measure tumor load, we quantified the number of cancer cells on the day of infection (day 0) and 6 days post-infection (PI) (day 6) by performing qPCR on DNA copies of the GFP gene, which was carried by the tumor cells but not the hosts (see [Experimental Procedures](#)). We chose to measure tumor load at 6 days PI to allow the cancer time to grow, but not so much time as to pass the median time to death for flies given high initial cancer doses. For each initial dose, cancer cells grew about 10-fold by day 6 PI in OR flies (Figures 1B and S1).

We generated a cancer tolerance curve by plotting median time to death for a given dose of cells against the cancer growth (i.e., the number of cells measured 6 days post-inoculation for that inoculation dose). (Figure 1C). These data were fit with a linear regression model ($r^2 > 0.94$) (Table S1). This design allows the health of these flies to be described with two parameters: The first is vigor (the health of the animal in the absence of disease, which in this case is around 30 days, and the second is the slope of the curve, which for this curve is -4.080 days per log of tumor load (Figure 1C).

Natural Variation of Cancer Resistance

To investigate how genetic variation might influence resistance and/or tolerance to cancer, we used two natural variant fly lines from the *Drosophila* Genetics Reference Panel (DGRP; lines

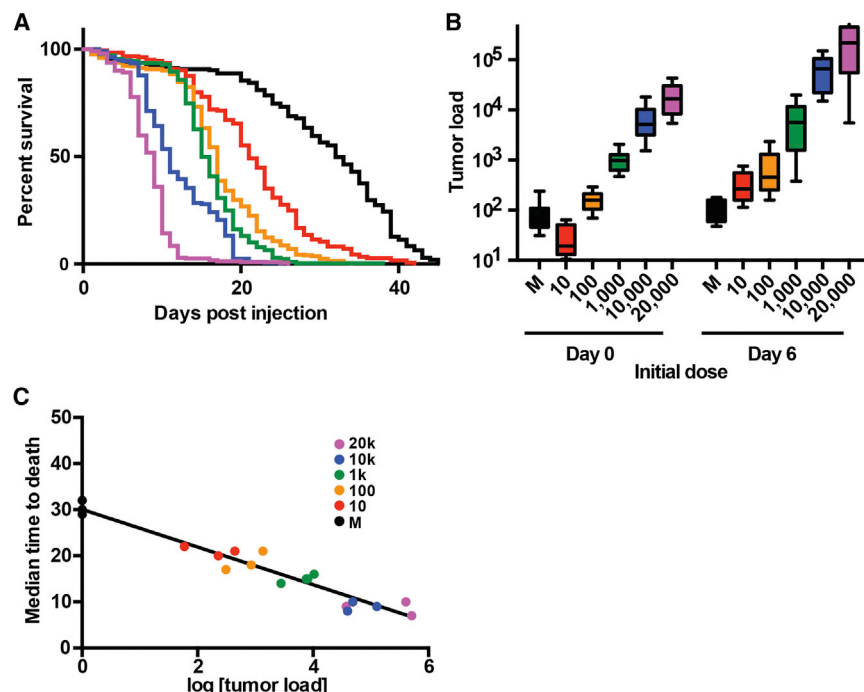


Figure 1. A Cancer Disease Tolerance Curve Establishes a Framework for Separating Resistance and Disease Tolerance to Cancer

Wild-type (Oregon-R) adult male flies were injected with doses varying from 10–20,000 K-Ras hyperplastic cancer cells and were monitored for survival (disease progression) and cancer load (elicitor load).

(A) Survival curves were monitored for flies injected with 10–20,000 K-Ras hyperplastic cancer cells ($n \geq 180$ flies per dose). The survival curves are significantly different (**** $p < 0.0001$, log-rank [Mantel-Cox] test).

(B) Ras^{v12}-H7 fly cancer cells were injected to adult flies. The initial dose (day 0) and subsequent cancer growth (day 6) were quantitatively measured using a *gfp* marker present in the cancer cells but not the flies ($n \geq 150$ flies per dose per day).

(C) A cancer disease tolerance curve was prepared by plotting pairs of cancer load and survival data for 18 cancer load/MTD pairs ($n \geq 110$ flies per data point). This curve was fit with a linear regression model ($r^2 > 0.94$) (Table S1).

RAL-358 and RAL-359) (Mackay et al., 2012). We selected RAL-358 and RAL-359 based on data from a pilot screen (Figure S2A). We generated a survival dose-response curve for these fly lines as described above (Figures S2B–S2G). RAL-358 died significantly faster than both wild-type and RAL-359 at all cell doses other than the highest dose of 20,000 cells (Figures 2A and S2B–S2G). To determine the role resistance played in these survival changes, we measured the tumor load of these lines at 6 days PI. RAL-358 had consistently higher loads than either wild-type or RAL-359 when injected with an initial dose of 10, 100, or 1,000 cells (Figures 2B and S1). All three fly lines had equal tumor loads 6 days PI when injected with high initial doses (10,000 or 20,000 cells) (Figures S1A and S1B). These data demonstrate that RAL-358 has a resistance defect, experiencing more than 100-fold tumor growth when given low initial doses of tumor cells (Figures 2B–2D and S1).

Resistance and disease tolerance are not mutually exclusive defense strategies but can act in concert in host defense. We wondered whether the decrease in survival in cancer-laden RAL-358 could also have a tolerance component. To investigate the differences in cancer disease tolerance, we plotted tolerance curves for all three fly lines (Figure 2C). We found that they had similar tolerance curves, suggesting that tolerance was not changing. Changes in resistance, however, are apparent in the growth plots and disease tolerance curves (Figures 2B–2D and S1). For example, the data for RAL-358 in the tolerance curve are shifted toward the bottom right of the plot compared to wild-type and RAL-359 (Figure 2C). The resistance defect we found in RAL-358 is dose dependent, manifesting only at low initial tumor inoculations (10–100 cells) (Figure 2D). This could be due to decreased immune surveillance sensitivity, where defenses, which can protect against tumor growth (Pastor-Pareja

et al., 2008), turn on only at higher initial tumor loads. These experiments highlight the importance of performing a dose-response curve when testing a health insult.

Applying Disease Tolerance to Cancer

Although the importance of resistance to cancer is well understood and has been well leveraged in current cancer treatment programs, cancer disease tolerance is not explicitly studied. There is evidence in some human cancers that health loss and survival rates do not always correlate with cancer burden, provocatively suggesting a role for cancer disease tolerance where hosts show different health effects given equal tumor loads (Heyneman et al., 2001; Patz et al., 2000). Our line of inquiry here provides a quantitative conceptual methodology for identifying unexplored aspects of host-cancer interactions that could be utilized to improve the treatment and outcomes for cancer patients, even when reduction or removal of cancer tissue is not possible.

EXPERIMENTAL PROCEDURES

Flies, Cells, and Media

The Oregon-R strain was used as a wild-type control, while *Drosophila* Genetic Reference Panel lines RAL-358 and RAL-359 were used as natural variants. Fly strains were obtained from the Bloomington *Drosophila* Stock Center (Mackay et al., 2012). Flies were kept in standard fly bottles containing dextrose medium (129.4 g dextrose, 7.4 g agar, 61.2 g corn meal, 32.4 g yeast, and 2.7 g tegosept per liter; polypropylene round-bottom, 8-oz bottles plugged with bonded dense weave cellulose acetate plugs, Genesee Scientific catalog no. 49-100) and were housed at 25°C with 65% relative humidity and a 12-hr-light and 12-hr-dark cycle. The hyperplastic cell line Ras^{v12}-H7, marked with a *UAS-GFP* construct, was obtained from the *Drosophila* Genomics Resource Center (Simcox et al., 2008). Cells were cultured as previously described (Simcox et al., 2008; Simcox, 2013). Briefly, cells were cultured in

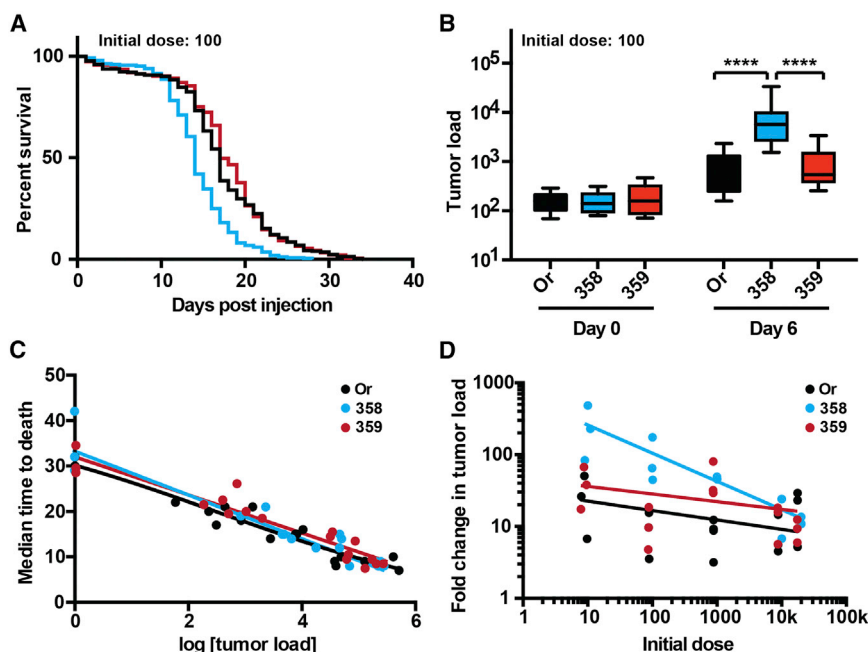


Figure 2. Genetic Variation Alters Resistance to Cancer

Wild-type (Oregon-R) and natural variant adult male flies were injected with doses varying from 10 to 20,000 KRas hyperplastic cancer cells and were monitored for survival (disease progression) and cancer load (pathogen load). Wild-type (Oregon-R) is in black, RAL-358 is in blue, and RAL-359 is shown in red.

(A) A survival curve of adult flies injected with 100 cancer cells, comparing wild-type and natural variant flies. RAL-358 dies significantly faster than wild-type or RAL-359 (**** $p < 0.0001$, log-rank [Mantel-Cox] test) ($n \geq 180$ flies per line), whereas there is no significant difference between wild-type and RAL-359.

(B) A cancer growth plot showing the initial dose of 100 cells and the cancer burden of flies 6 days post-injection. RAL-358 has a significantly higher cancer load than either wild-type or RAL-359 (**** $p < 0.0001$, two-way ANOVA Tukey's multiple comparisons test) ($n \geq 150$ flies per dose per day). (C) A cancer disease tolerance curve was prepared for each of the three fly lines (wild-type, RAL-358, and RAL-359) by plotting pairs of cancer load and survival data for 18 cancer load/MTD pairs for each line ($n \geq 110$ flies per data point).

These curves were each fit with a linear regression model ($r^2 > 0.94$ for wild-type, $r^2 > 0.91$ for RAL-358, and $r^2 > 0.91$ for RAL-359). The slope of these lines is similar (-4.1 for wild-type, -4.8 for RAL-358, and -4.2 for RAL-359) and the 95% confidence intervals overlap.

(D) The ratio of cancer growth over 6 days PI. Wild-type is in black, RAL-358 is in blue, and RAL-359 is shown in red ($n \geq 150$ flies per line per day in all of these experiments). The data for each of these lines were fit with a log-log non-linear regression, 0.12 , 0.57 , and 0.13 R^2 for Or, RAL-358, and RAL-359, respectively. The slope of the log-log non-linear regression is -0.13 , -0.4 , and -0.11 for Or, RAL-358, and RAL-359, respectively. The 95% confidence interval for the slope of RAL-358 remains negative, while the 95% confidence interval for the slope of wild-type and RAL-359 ranges from negative to positive. In a non-linear log-log line regression F test, one curve does not fit all the data ($p < 0.0001$).

Schneider's media supplemented with 10% heat-inactivated fetal calf serum (Sigma catalog no. F4135-100ML) and 1% penicillin-streptomycin solution (HyClone catalog no. SV30010). Ras^{V12}-H7 cells are strongly adherent and were passaged by first rinsing the cells in chilled PBS and then incubated for 5 min in 0.05% trypsin (HyClone catalog no. SH30236.01). Cells were collected and diluted into an equal volume of media, centrifuged for 2 min, and resuspended in new media. For infection experiments, the cells were quantified using a hemocytometer and diluted to the desired doses. Frozen stocks of cells were produced as previously described (Simcox, 2013).

Cell Injections into Adults

Different doses of Ras^{V12}-H7 cells (10, 100, 1,000, 10,000, and 20,000 cells) were injected into adult male flies aged 5–7 days old with control flies being injected with cell-culture media. Flies received 50-nl injections in the anterior abdomen. Injections were performed using a Picospritzer III (Parker Instrumentation) and a pulled glass needle as previously described (Ayres et al., 2008). Cancer cell doses were counted using a hemocytometer immediately prior to injection. Each dose was injected into ≥ 60 adult male flies, and each experiment was replicated three times, thus $n = \geq 180$ flies per dose, per fly line tested, resulting in $\geq 3,240$ individual *D. melanogaster* being injected for the survival portion of these experiments. Survival of the cancer injected flies was counted every 24 hr post-infection (PI) until all flies were dead. The median time to death (MTD) for each experiment was determined as used as a measure of disease progression.

Quantifying Cancer Cell Growth

Ras^{V12}-H7 cells carry an integrated UAS-GFP marker. Cancer load in flies was determined by qPCR, amplifying DNA copies of the *gfp* marker using previously published primers (Simcox et al., 2008; Portugal et al., 2011). Total DNA was extracted using phenol-chloroform. We quantified *gfp* by pooling five adult male flies to make one sample, and ten samples were taken per

dose both on the day of infection (day 0) and 6 days PI (day 6). Thus, $n \geq 50$ flies per dose per day, for five doses plus media controls, and each experiment was replicated three times, resulting in the extraction of $\geq 1,800$ flies in batches of five. qPCR was performed on a StepOnePlus qRT-PCR system and analyzed using StepOne software v.2.2.2 (Applied Biosystems). Reactions were done using SYBR Green PCR Master Mix (Applied Biosystems) using 15- μ l reactions, in 96-well plates, using a relative standard curve. Following a primary denaturation of 10 min at 95°C, the reactions were done for 40 cycles of 95°C for 15 s and 60°C for 1 min. We used *gfp* primers from Portugal et al. (2011) 5'-GTC AGT GGA GAG GGT GAA GG-3' and 5'-ACT TCA GCA CGT GTC TTG TAG TTC-3'. The standard curve was performed using a serial dilution series (1:10) of Ras^{V12}-H7 cells, quantified using a hemocytometer. The *gfp* abundance relative to our standard curve provided a quantitative measure of initial cancer dose and cancer burden after 6 days post-infection. Using MTD as a measure of disease progression and the cell growth at 6 days PI, we were able to plot disease tolerance curves as shown in Figures 1 and 2.

Statistical Analyses

Statistical analyses were done using GraphPad Prism version 6 for Mac OS X. The log-rank (Mantel-Cox) test was used to evaluate the survival curves. Two-way ANOVA with Tukey's multiple comparisons tests were used to analyze cancer growth among the fly lines. Disease tolerance curves were fit with a linear regression model after testing several other models (four-parameter sigmoid, three-parameter sigmoid, and quadratic).

SUPPLEMENTAL INFORMATION

Supplemental Information includes two figures and one table and can be found with this article online at <http://dx.doi.org/10.1016/j.celrep.2015.09.052>.

AUTHOR CONTRIBUTIONS

A.R.D. and D.S.S. conceived and designed the experiments. A.R.D. performed the experiments. A.R.D. and D.S.S. analyzed and interpreted the data and wrote and revised the manuscript.

ACKNOWLEDGMENTS

We would like to thank lab members and colleagues for critical reading of the manuscript (Alejandra Hotson, Alexander Louie, Brenda Torres, Damian Trujillo, Jose Oliveira, Katherine Cumnock, and Poonam Rath). We also thank Alejandra Hotson, Alexander Louie, Jose Oliveira, Katherine Cumnock, and Poonam Rath for their help in counting fly survival. We thank Amanda Simcox, the Bloomington *Drosophila* Stock Center (NIH P40OD018537) and the *Drosophila* Genomics Resource Center (NIH 2P40OD010949-10A1) for fly stocks and cell cultures. A.R.D. is a Howard Hughes Medical Institute fellow of the Life Sciences Research Foundation. This work was supported by an NIH Pioneer Award (DP1 AT007753) to D.S.S. and an NIH training grant (5T32HG000044-17) to A.R.D.

Received: December 15, 2014

Revised: June 3, 2015

Accepted: September 18, 2015

Published: October 22, 2015

REFERENCES

- Ayres, J.S., and Schneider, D.S. (2009). The role of anorexia in resistance and tolerance to infections in *Drosophila*. *PLoS Biol.* 7, e1000150.
- Ayres, J.S., Freitag, N., and Schneider, D.S. (2008). Identification of *Drosophila* mutants altering defense of and endurance to *Listeria monocytogenes* infection. *Genetics* 178, 1807–1815.
- Caldwell, R.M., Schafer, J.F., Compton, L.E., and Patterson, F.L. (1958). Tolerance to cereal leaf rusts. *Science* 128, 714–715.
- Heyneman, L.E., Herndon, J.E., Goodman, P.C., and Patz, E.F., Jr. (2001). Stage distribution in patients with a small (< or = 3 cm) primary nonsmall cell lung carcinoma. Implication for lung carcinoma screening. *Cancer* 92, 3051–3055.
- Howick, V.M., and Lazzaro, B.P. (2014). Genotype and diet shape resistance and tolerance across distinct phases of bacterial infection. *BMC Evol. Biol.* 14, 56.
- Iwasaki, A., and Pillai, P.S. (2014). Innate immunity to influenza virus infection. *Nat. Rev. Immunol.* 14, 315–328.
- Mackay, T.F., Richards, S., Stone, E.A., Barbadilla, A., Ayroles, J.F., Zhu, D., Casillas, S., Han, Y., Magwire, M.M., Cridland, J.M., et al. (2012). The *Drosophila melanogaster* Genetic Reference Panel. *Nature* 482, 173–178.
- Medzhitov, R., Schneider, D.S., and Soares, M.P. (2012). Disease tolerance as a defense strategy. *Science* 335, 936–941.
- Pastor-Pareja, J.C., Wu, M., and Xu, T. (2008). An innate immune response of blood cells to tumors and tissue damage in *Drosophila*. *Dis. Model. Mech.* 1, 144–154, discussion 153.
- Patz, E.F., Jr., Rossi, S., Harpole, D.H., Jr., Herndon, J.E., and Goodman, P.C. (2000). Correlation of tumor size and survival in patients with stage IA non-small cell lung cancer. *Chest* 117, 1568–1571.
- Portugal, S., Carret, C., Recker, M., Armitage, A.E., Gonçalves, L.A., Epiphany, S., Sullivan, D., Roy, C., Newbold, C.I., Drakesmith, H., and Mota, M.M. (2011). Host-mediated regulation of superinfection in malaria. *Nat. Med.* 17, 732–737.
- Råberg, L. (2014). How to live with the enemy: understanding tolerance to parasites. *PLoS Biol.* 12, e1001989.
- Råberg, L., Sim, D., and Read, A.F. (2007). Disentangling genetic variation for resistance and tolerance to infectious diseases in animals. *Science* 318, 812–814.
- Regoes, R.R., McLaren, P.J., Battegay, M., Bernasconi, E., Calmy, A., Günthard, H.F., Hoffmann, M., Rauch, A., Telenti, A., and Fellay, J.; Swiss HIV Cohort Study (2014). Disentangling human tolerance and resistance against HIV. *PLoS Biol.* 12, e1001951.
- Rose, P.P., Hanna, S.L., Spiridigliozzi, A., Wannissorn, N., Beiting, D.P., Ross, S.R., Hardy, R.W., Bambina, S.A., Heise, M.T., and Cherry, S. (2011). Natural resistance-associated macrophage protein is a cellular receptor for sindbis virus in both insect and mammalian hosts. *Cell Host Microbe* 10, 97–104.
- Rottschaefer, S.M., and Lazzaro, B.P. (2012). No effect of *Wolbachia* on resistance to intracellular infection by pathogenic bacteria in *Drosophila melanogaster*. *PLoS ONE* 7, e40500.
- Schafer, J.F. (1971). Tolerance to plant disease. *Annu. Rev. Phytopathol.* 9, 235–252.
- Simcox, A. (2013). Progress towards *Drosophila* epithelial cell culture. In *Epithelial Cell Culture Protocols*, Second Edition, S.H. Randell and M.L. Fulcher, eds. (Humana Press), pp. 1–11.
- Simcox, A., Mitra, S., Truesdell, S., Paul, L., Chen, T., Butchar, J.P., and Justiniano, S. (2008). Efficient genetic method for establishing *Drosophila* cell lines unlocks the potential to create lines of specific genotypes. *PLoS Genet.* 4, e1000142.
- Vale, P.F., Fenton, A., and Brown, S.P. (2014). Limiting damage during infection: lessons from infection tolerance for novel therapeutics. *PLoS Biol.* 12, e1001769.